

Effects of Cisplatin on the Sperm Morphology of *Rattus Orvegicus*

Abstract: The mutagenicity and genotoxicity of chemotherapeutic drugs used as chemical antibiotics in most tumours as anticarcinogenic agents were studied. Cis-diammine-dichloroplatinum (cisplatin) is a commonly used chemotherapeutic agent in the treatment of ovarian, testicular and bladder cancer. It is used intravenously, intraarterially and intraperitoneally. Group I adult male rats were injected with a single dose at a level of $1/2$ LD₅₀ (16 mg/kg body weight (a half carcinogenic dose). Group II rats were treated with 9 mg/kg body weight (recommended dose) intraperitoneally injected. The abnormally shaped sperms were recorded after 1, 2, 3, 4 and 5 weeks post-injection. The frequency of abnormally shaped sperms was noted within the first three weeks post-injection, and highly significant abnormalities were recorded at the end of second week (181.4 ± 24.9) and (145.1 ± 28.6) in Groups I and II, respectively. Recovery from the genotoxicity of cisplatin was achieved at the end of the 5th week.

Key words: Cisplatin, Sperm morphology, *Rattus orvegicus*.

Introduction

Cisplatin is of value in the treatment of metastatic tumours of the testis, usually as a major component of combination chemotherapy regimens, and particularly in combination with bleomycin and etoposide. It is also used in metastatic ovarian tumours and advanced bladder cancer, and has been reported to be active against a wide range of other solid tumours (Shernian *et al.* 1985).

The cytotoxic effect of cisplatin is thought to result from the covalent binding of the cisplatin to DNA. The resultant adducts are cross-links of three

تأثير السيزبلاطين على أشكال الحيوانات المنوية للجرذان البالغة (راتس نورفيجيكس)

فوزية عبد الهادي زايد

المستخلص: استهدف هذا البحث تقنين الآثار الجانبية للعقار الكيميائي سيزبلاطين على أشكال الحيوانات المنوية للجرذان البالغة. استخدم في هذا البحث 60 جرذ ذكر بالغ متوسط وزنه من 100-120 جرام وقسمت إلى مجموعتين أساسيتين. 30 جرذ في كل مجموعة. وقسمت كل مجموعة إلى 6 مجموعات صغيرة (خمس جرذان لكل مجموعة). أعتبرت المجموعة الفرعية الأولى في كل مجموعة كبيرة مجموعة ضابطة عوملت بمحلول فسيولوجي، بالحقن في التجويف البريتوني. عوملت جرذان المجموعة الأولى بجرعة نصف مسرطنة تعادل نصف الجرعة نصف المميتة LD₅₀ (16 ملليجرام لكل كيلو جرام من وزن الجسم) بالحقن مرة واحدة في التجويف البريتوني ذبحت الجرذان بعد 1, 2, 3, 4, 5 أسابيع على التوالي. بينما عوملت جرذان المجموعة الثانية بالجرعة الأمانة (9 ملليجرام لكل كيلو جرام من وزن الجسم) وتم ذبح الجرذان بعد 1, 2, 3, 4, 5 أسابيع على التوالي. تم أخذ الخصية وتشريحها ووضع البربخ العلوي في محلول ملحي ثم عمل مسحة منوية على شرائح نظيفة وصبغت الشرائح بصبغة الفولجين النووية. وتم الفحص بالميكروسكوب الضوئي بعدسة عينة قوتها 10x وعدسة زينية قوتها 100x بمعدل 1000 حيوان منوي لكل جرذ.

وقد تبين من الدراسة أن المجموعات المعالجة بواسطة الجرعة الأمانة والجرعة نصف المسرطنة أدت إلى زيادة ذات دلالة إحصائية في عدد الحيوانات المنوية المشوهة خلال الأسابيع الثلاثة الأولى بعد الحقن. وكان أقصى معدل للتشوه الشكلي للحيوانات المنوية بعد الأسبوع الثاني من الحقن بمعدل 148.2%، 180.4% لكل من الحيوانات المعالجة بالجرعتين المختلفتين على التوالي إذا ما قورنت بالمجموعة الضابطة 48.9%. وتم الاستشفاء من الآثار الجانبية للسيزبلاطين في الأسبوع الرابع والخامس. من هذا البحث يلاحظ أن استخدام السيزبلاطين في جرعة الأمانة يؤدي إلى أعراض جانبية ممثلة في تشوهات في الحيوانات المنوية مما يقلل من درجة الخصوبة.

كلمات مدخلية: السيزبلاطين، الحيوانات المنوية، الجرذان البالغة

Fawzia A Zaied
Zoology Dept, Faculty of Science, Zagazig University
Egypt
Tel. No: 055 2300818, 055 2360818
Fax No: 055 2303252

types: intrastrand, interstrand and DNA-protein (Sherman & Lippard, 1987). The most common adduct is the intrastrand cross-link bridging adjacent guanines at the N-7-position (Pinto & Lippard, 1985 and Eastman, 1987). It also inhibits DNA synthesis and replication by inactivating the template.

Other observations, related to the genotoxic effects of cisplatin therapy include damage to microtubule formation (Jnoue *et al.* 1985) and G2 arrest (Andrews & Howell, 1990). Most studies of the genotoxicity of cisplatin on germ cells have been done in males. However, male and female germ cells differ in their susceptibility to mutation induction. Thus, genotoxic data extrapolated to female germ cells must be treated with caution. (Jekunen *et al.* 1994). The present study is an attempt to explore if cisplatin therapy may or may not affect sperm shape of male rats.

Material and Methods

Cis-diammino dichloro platinum II (cisplatin):

Cisplatin is an inorganic platinum-containing complex. It is thought to act as an alkylating agent which is potentially carcinogenic. Its antineoplastic action is cell cycle nonspecific (Ogawa, 1985). Cisplatin was synthesized by Marceils and Reedijk (1985). It is a yellow powder, slightly soluble in water and in 0.9 sodium chloride solution (1 mg/1 ml of saline solution) and must be stored in airtight containers and protected from light. Cisplatin is administered intraperitoneally and the therapeutic dose for humans is 100 mg/Ms day as a single dose monthly (Vermorken, *et al.* 1984). The recommended dose for rats is 9 mg/kg body weight as a single dose (Adler, & El-Tarras, 1990), and $1/2$ LD₅₀ of cisplatin in male rats is 16mg/kg body weight, which is half a carcinogenic dose (Connors, *et al.* 1982).

In the present work 60 adult male rats (*Rattus norvegicus*) were obtained from the farm of the Faculty of Veterinary Medicine, Zagazig University, weighing an average of 100 ± 20 gm. The specimens were classified into 2 main groups, each of 30 rats. Each group was subdivided into 6 subgroups each of 5 rats. In both groups, rats of the first subgroup were given 1.0 ml saline solution and served as a control group. Subgroups 2-6 represented rats of weeks 1-5 respectively.

Group I

Each rat in the 5 subgroups received an evening intraperitoneal injection of a single dose of $1/2$ LD₅₀ cisplatin (1.6 mg/rat/day). Five rats were sacrificed after 1, 2, 3, 4 and 5 weeks and sperm smears were taken from the caudal epididymis of the testes.

Group II

Each rat from 25 intact rats was intraperitoneally injected with the recommended dose of 0.9 mg/rat/day as a single dose in the evening. After 1, 2, 3, 4 and 5 weeks, five rats were sacrificed and sperm smears were taken from the caudal epididymis of the testes. Sperm smears of 60 rats were stained by Feulgen nuclear stain (Feulgen & Rossenbeck, 1924), and 1000 sperms were microscopically examined for each rat. A binocular microscope, with x10 eye piece and x100 oil immersion objective lenses was used. Abnormally shaped sperms were recorded. Micrographs were taken whenever necessary. Statistical analysis for each treatment was evaluated by a dispersion test based on χ^2 (Snedecor & Cochran, 1976).

Results

The rat sperm consists of two main parts, the head and the tail. The head is triangular in shape, having a broad base and pointed tip. The pointed tip has a characteristic curvature. The head region is mostly occupied by the nucleus. The tail is several times the length of the head. It begins as a cylindrical part at the base of the head region and then the diameter decreases progressively and ends in a filamentous form (see Fig. 3).

In the present study, the frequency of normal sperm shape reached 951.1 ± 9.3 /1000 sperms. Occasionally, some sperm cells may be present with rather an unusual shape. However, such cells appear in limited numbers with an average of 48.9 ± 9.3 /1000 sperms. Cisplatin in a dose of 0.9 mg/rat/day intraperitoneally injected into rats resulted in an obvious effect on the sperm shape when compared with the control group. The abnormally shaped sperms were significantly increased after the first and second weeks post-injection. This increase persisted for 4 weeks post-injection with a peak after the second week (Tables 1 & 2).

Thereafter, there was a gradual decline in frequency that continued until the end of the experiment (5 weeks). However, there was always a variation in the response between rats treated with the different doses of cisplatin. The higher response

was reported after the second week, reaching 148.2 ± 27.6 and 180.4 ± 24.9 l in rats treated with the recommended dose and rats treated with $1/2$ the carcinogenic dose of cisplatin, respectively. By the end of the fourth week, the average number of deformed sperms reached 67.7 ± 14.8 and 53.4 ± 11.5 in both groups of treated rats, respectively, which was within the control values. However, the abnormally shaped sperms remained higher in number than the control level at the end of the 5th

week. After the 5th week, the frequency returned to its normality.

The sperm shape abnormalities involved either the head or the tail. In many cases, both the head and tail were deformed (Tables 1 & 2).

The majority of deformed sperm abnormalities involved the head morphology, which reached a peak in week two post-injection, reaching 4.6% and 7.4% in both groups of treated rats respectively compared with the control of 2.4%, as in Table (3) and Fig. (2).

Table 1: Effect of cisplatin given intraperitoneally in a dose of (0.9 mg/rat/day) on the sperm morphology of rats.

Weeks	Normal sperms /1000 M \pm SD	Deformed sperms / 1000 M \pm SD			Total abnormal sperms/1000 M \pm SD
		Head	Tail	Head and tail	
Control	951.1 \pm 9.3	24.1 \pm 4.7	10.3 \pm 2.3	14.5 \pm 2.6	48.9 \pm 9.3
1	860.7 \pm 24.1	33.7 \pm 2.6	16.3 \pm 7.3	89.3 \pm 14.2***	139.3 \pm 24.1***
2	851.8 \pm 27.6	49.5 \pm 10.1**	38.4 \pm 8.0**	60.3 \pm 9.5**	148.2 \pm 27.6***
3	866.2 \pm 28.7	61.5 \pm 16.2**	24.2 \pm 7.1*	48.1 \pm 5.4**	133.8 \pm 28.7***
4	932 \pm 14.8	23.8 \pm 5.2	21.0 \pm 6.3	22.2 \pm 3.3*	67 \pm 14.8
5	950.1 \pm 11.8	20.5 \pm 3.2	14.5 \pm 2.5	14.9 \pm 6.1	49.9 \pm 11.8

* P<0.05,

** P < 0.01,

*** P < 0.001

Table 2: Effect of intraperitoneal injection of cisplatin (1.6 mg/rat/day) on the sperm morphology of rats.

Weeks	Normal sperms /1000 M \pm SD	Deformed sperms / 1000 M \pm SD			Total abnormal sperms/1000 M \pm SD
		Head	Tail	Head and tail	
Control	951.1 \pm 9.3	24.1 \pm 4.4	10.3 \pm 2.3	14.5 \pm 2.6	48.9 \pm 9.3
1	840.3 \pm 25.5	85.3 \pm 14.3**	20.2 \pm 3.1**	54.4 \pm 8.1**	159.7 \pm 25.5***
2	819.6 \pm 24.9	90.4 \pm 14.5**	21.3 \pm 2.1**	68.7 \pm 8.3***	180.4 \pm 24.9***
3	866.2 \pm 28.7	71.5 \pm 16.2**	14.1 \pm 5.3	48.2 \pm 7.2**	133.8 \pm 28.7***
4	946.6 \pm 11.5	29.0 \pm 6.7	5.3 \pm 0.9	19.1 \pm 3.9	53.4 \pm 11.5
5	960.5 \pm 12.5	20.3 \pm 8.2	5.0 \pm 1.3	14.2 \pm 3.0	39.5 \pm 12.5

* P<0.05,

** P < 0.01,

*** P < 0.001

Table 3: Comparison of % abnormality of sperm heads in the rats treated with both doses of cisplatin.

Weeks	Deformed head / 6000 sperms											
	Rats treated with 0.9 mg/rat/day						Rats treated with $1/2$ LD ₅₀ 1.6 mg/rat/day					
	Hooks		Unusual head		Total		Hooks		Unusual head		Total	
	N	%	N	%	N	%	N	%	N	%	N	%
Control	58	1.0	86	1.4	144	2.4	-	-	-	-	-	-
1	61	1.2	141	2.4	202	3.4	130	2.1	281	4.7	411	6.8
2	65	1.2	211	3.5	276	4.6	192	3.2	250	4.2	442	7.4
3	48	0.8	138	2.3	186	3.1	137	2.3	192	3.2	329	5.5
4	43	0.8	95	1.7	138	2.5	95	1.7	39	1.6	134	2.3
5	41	0.7	85	1.4	126	2.1	39	1.3	81	1.7	120	2.0

N = number of abnormal heads

The head deformities are presented in various forms. Sometimes the apical part showed an unusual curvature, or was abnormally straight and in many cases the size of the head was within the normal limits. In other cases, the size was abnormally large as it had an irregular outline which was rather difficult to describe, as in Fig. (3).

Abnormalities in the tail region varied between being duplicated at its junction with the head and other deformities in the tail, as in Figure (3). All of these deformities were mostly seen within three weeks post-injection of cisplatin, and also during the recovery from the side effects of cisplatin, mostly in rats treated with the recommended dose but less than with those rats treated with the 1/2 carcinogenic dose.

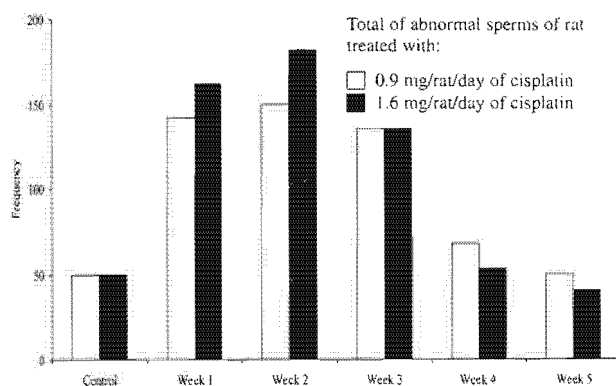


Fig. 1: The frequency of abnormally shaped sperm heads of the rats treated with both doses of cisplatin.

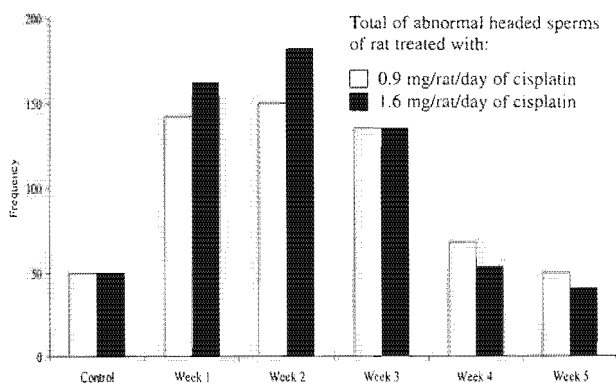


Fig. 2: Comparison between the percentage of abnormal heads of sperm heads of rats treated with both doses of cisplatin.

Discussion

In an attempt to find out the way in which cisplatin affects germ cell development, it was found that the therapeutic dose (9 mg/kg body weight) resulted in an abundance of abnormally shaped sperm heads. The peak of abnormality was observed in the second week post-injection. Abnormalities in shape affected the head region, tail region or both.

Deformities of the tail most probably resulted in improper motility, which in turn limited the chance for such a sperm to fuse with an ovum. In our opinion the deformities of the tail region indicated that the cytotoxicity of cisplatin has a high level of adduct formation with mitochondrial DNA, where the tail formation and motility of sperms depend on the energy provided from mitochondria. This notion is in agreement with the results recorded by Adrie *et al.* (1985), Plooy *et al.* (1985), Andrews *et al.* (1988) and Sabet *et al.* (1990). They attributed increased sensitivity of germ cells to cisplatin and high toxicity of cisplatin to intra-lysosomal storage or metabolites. This means that lethality should be due to cytoplasmic effects of the drug and high level of adduct formation with mitochondrial DNA, which could occur and cause dysfunction of the cells. The head shape abnormalities are markedly increased. One type of head shape abnormality affected its tapering anterior part; such sperms with untapered anterior region might have been also unable to fuse with ovum for zygote formation. In addition, there are various head shapes that lead to an irregular form which was rather difficult to describe.

The present results point to the sensitivity of the component of head region to cisplatin. In most cases the nucleus occupies all the head region, and the genotoxicity of cisplatin has formed intrastrand cross-links with nuclear DNA or with defective repair. This suggestion is supported by the observations of many previous authors (Roberts *et al.* 1982) and (Plooy *et al.* 1984). It is postulated that potentially lethal platinum DNA adduct appears to be the intrastrand cross-link. This is because it amounts to more than 1% of the total amount of platinum that had reacted with nuclear DNA, and because of high penetration of cisplatin into the nucleus and the tendency of nuclear germ cells to accumulate platinum. Other authors, such as Roberts and Friedlas, (1987) Adler and El-Tarras (1989 & 1990) and Zamble *et al.* (1996) have found that cisplatin produces a clastogenic effect on differentiated mouse spermatogenic cells with a number of aberrations per cell that increase in a dose-dependent manner. Cisplatin also induced genetic aberration during the pre-meiotic phase of DNA synthesis (Adler & El-Tarras 1990) and (Kenneth *et al.* 1999).

More sperms became affected at the end of the second week post-injection with cisplatin. We suggest that late spermatids seem to be the most sensitive cells in the spermatogenic series to cisplatin with longer time needed for recovery from cisplatin genotoxicity, indicating that other cell

stages in spermatogenesis can also be affected by cisplatin. This assumption is supported by many authors. Meistrich *et al.* (1982) found that differentiated murine spermatogonia, spermatocytes and spermatids were all sensitive to cellular death after cisplatin therapy while stem cells were relatively resistant. Genesca *et al.* (1990) noted an increase in chromosome breaks and univalents in all male germ cell tested with cisplatin treatment. Drasga *et al.* (1983), Johnson *et al.* (1984), Brenner *et al.* (1985) and Fossa *et al.* (1985) revealed that most patients with testicular malignant tumors are rendered azospermic by cisplatin therapy and restoration of spermatogenesis occurs in many patients after cessation of therapy, but not all patients will demonstrate return of fertility.

In the present study, the recovery from side effects of platinum extended for 5 weeks or more in male rats treated with cisplatin until it reached the control limit, showing the tendency of platinum to form DNA adduct strands and accumulate with nuclear DNA. Our results coincide with the data obtained by Rothmann and Weick (1981) and Guichun *et al.* (1999). They reported that most patients with testicular cancer recover by day 39 post treatment with cisplatin. Schwartz & Vidone (1981), Ward *et al.* (1982) and Gershenson *et al.* (1983) suggested that the incidence of germ cell failure is related to the age and period at which the patients received cisplatin therapy.

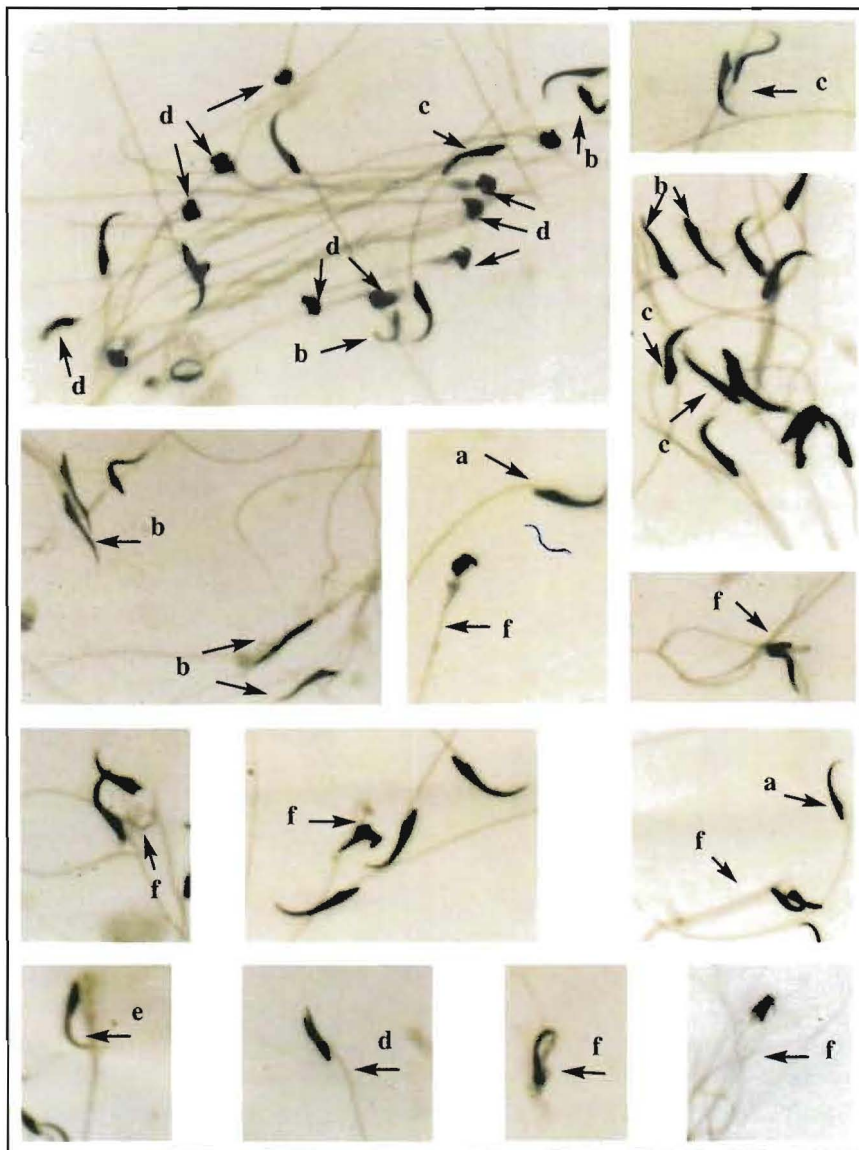


Fig. 3: Sperm cells showing:

- a) Normal sperm, b) Sperms showing unusual acute curvature, c) Sperms with unusual hooks
- d) Sperms with unusual heads, e) Sperms with unusual tails, and f) Sperms with unusual heads and tails

References

- Adler, L. and El-Flirras, A.** (1989) Clastogenic effects of Cis-diamminedichloroplatinum. I. Induction of chromosomal aberrations in somatic and germinal cells of mice. *Mutat. Res.* **211**: 131-137.
- Adler, L. and El-Tarras, A.** (1990) Clastogenic effects of Cis-diamminedichloroplatinum. II. Induction of chromosomal aberrations in primary spermatocytes and spermatogonial stem cells of mice. *Mutat. Res.* **243**: 173-178.
- Adrie, C. M., Pooy M. and Pul H. M.** (1985) Formation and repair of DNA inter strand cross-links in relation to cytotoxicity and unscheduled DNA synthesis, induced in control and mutant human cells treated with Cisplatin. *Cancer Res.* **45**: 4178-4184.
- Andrews, P. A. and Howell, S. B.** (1990) Cellular pharmacology of Cisplatin. Perspectives on mechanisms of acquired resistance. *Cancer Cells.* **2**: 35-43.
- Andrews, P. A., Velury, S., Mann, S. C. and Howell, S.B.** (1988) Cis-diammine-dichloroplatinum (II) accumulation in sensitive and resistant human ovarian carcinoma cells. *Cancer Res.* **45**: 68-73.
- Brenner, J., Vogrin, D. and Whitmore, W.** (1985) Effect of treatment on fertility and sexual function in males with metastatic nonseminomatous germ cell tumors of testis. *Am. J. Clin. Oncol.* **8**: 178-182.
- Connors, T., Jones, M., Ross, W., Braddock, P., Khokhar, A. and Tobe, M.** (1982) A new platinum complex with antitumour activity. *Chem. Bid. Interact.* **5**: 415-424.
- Drasga, R., Einhorn, L., Williams, S., Patel, D. and Stevens, E.** (1983) Fertility after chemotherapy for testicular cancer. *J. Clin. Oncol.* **1**: 179-183.
- Eastman, A.** (1987) Cross-linking of glutathione DNA by cancer chemotherapeutic platinum coordination complexes. *Chem. Bio. Interact.* **61**: 241-248.
- Feulgen, R. and Rossenbeck, H.** (1924) Mikroskopisch-chemischer Nachweis einer Nucleinsäure von Typus der Thymonud-einsäure und die darauf beruhende selektive Färbung von Zellkern in mikroskopischen Präparation. *Z. Physiol-Chem.* **26**: 136-203.
- Fossa, S., Ous, S., Abyholm, T., Norman, N. and Loeb, M.** (1985) Post-treatment fertility in patients with testicular cancer. Influence of Cisplatin II-based combination chemotherapy and of retroperitoneal surgery on hormone and sperm cell production. *Br. J. Urol.* **57**: 210-214.
- Genesca, A., Miro, R., Caballn, M., Benet J., Germa, and Egozcue, J.** (1990) Sperm chromosome studies in individuals treated for testicular cancer. *Hum. Reprod.* **5**: 286-290.
- Gershenson, D. M., Del Junco, G., Herson, J. and Rutledge, F. N.** (1983) Endodermal sinus tumour of the ovary: the M.D. Anderson experience. *Obstet Gynecol.* **61**: 194-202.
- Guichun Wang, L. M., Hallberg, E. W. and Englander, R.** (1999) Rapid SINE-mediated detection of cisplatin: DNA adduct formation in vitro and in vivo in blood. *Mutation. Res.* **434**: 67-74.
- Inoue, K., Mukaiyama, T., Mitsui U. and Ogawa, M.** (1985) In vitro evaluation of anticancer drugs in relation to development of drug resistance I the human tumor carcinogenic assay. *Cancer Chemother. Pharmacol.* **15**: 208-213.
- Jekunen, A. P., Hom, D. K., Alcaraz, J. E., Eastman, A. and Howell, S. B.** (1994) Cellular pharmacology of dischloro (ethylenediamine) platinum (II) in Cisplatin-sensitive and resistant human ovarian carcinoma cells. *Cancer Res.* **54**: 2680-2687.
- Johnson, D., Hainsworth, J., Linde, R. and Greco, F.** (1984) Testicular function following combination chemotherapy with Cisplatin, vinblastine, and bleomycin. *Med. Pediatr. Oncol.* **12**: 233-238.
- Kenneth, F., Grossmann, J.C., Brown, R.E. and Moses, E.** (1999) Cisplatin DNA cross-links do not inhibit S-phase and cause only a G2/M arrest in *Saccharomyces cerevisiae*. *Mutation. Res.* **434**: 29-39.
- Marceils, A. T. M. and Reedijk, J.** (1985) Binding of platinum compounds to nucleic acids with respect to the anti-tumor activity of Cisdiammine-dichloroplatinum (II) (cisplatin). II. Comparison of results from alkaline elution, DNA renaturation and DNA sedimentation studies. *Biochem. Biophys. Acta.* **655**: 152-166.
- Meistrich, M., Finch, M., DaCunha, F., Hacker, U. and Au, W.** (1982) Damaging effects of fourteen chemotherapeutic drugs on mouse testis cells. *Cancer Res.* **42**: 122-131.
- Ogawa, G. S.** (1985) Dispensing-pin problems. *Am J. Hosp. Pharm.* **42**: 1042-5.
- Pinto, A. L. and Lippard, S. J.** (1985) Binding of the anti-tumor drug Cisdiammine-dichloro-platinum (II) (cisplatin) to DNA. *Biochem. Biophys. Acta.* **780**: 167-180.
- Plooy, A., Van Dijk, M. and Lohman, P.** (1984) Induction and repair of DNA cross-links in Chinese hamster ovary cells treated with various platinum coordination compound with relation to platinum binding to DNA, cytotoxicity, mutagenicity, and antitumor activity. *Cancer Res.* **44**: 2043-2051.
- Plooy, A. C. M., Van Dijk, M., Berends, F. and Lohman, P. H. M.** (1985) Formation and repair of interstrand cross-links in relation to cytotoxicity and unscheduled DNA synthesis induced in control and mutant human cells treated with Cis-diamine-dichloroplatinum (II). *Cancer Res.* **45**: 4178-4184.
- Roberts, J. and Friedlos, F.** (1987) Quantitative estimation of Cisplatin induced DNA interstrand cross-links and their repair in mammalian cells: Relationship to toxicity. *Pharmacol Ther.* **34**: 215-246.
- Roberts, J., Pera, M. F., and Rawlings, C.J.** (1982) The role of DNA repair in the recovery of mammalian cells from cis-diamminedichloroplatinum (II) (cisplatin) induced DNA damage. *In: Natarajan, A.T. Obe, G. and Altmann H.* (eds.) *DNA Repair*;

- Chromosome Alterations and Chromatin Structure*. Elsevier North-Holland Biomedical Press, Amsterdam, pp. 223-246.
- Rothmann, S. A. and Weick, J. K.** (1981) Cisplatin toxicity for erythroid precursors. *N Engl J. Med.* **304**: 360.
- Sabet, M. K., Lu, Y., Leong, L., Haedick, K. and Scalon, K. J.** (1990) Differential oncogene amplification in tumor cells from a patient treated with Cisplatin and 5-fluorouracil. *Eur. J. Cancer* **26**: 383-390.
- Schwartz, P. E. and Vidone, R. A.** (1981) Pregnancy following combination chemotherapy for mixed germ cell tumour of the ovary. *Gynecol Oncol.* **12**: 373-378.
- Sherman, E. S., Gibson, D., Whang, A. H. and Lippard, S. J.** (1985) X-ray structure of the major adduct of the anticancer drug Cisplatin with DNA: Cis [Pt (NH₃)₂ {d (pGpG) }]. *Science* **230**: 412-417.
- Sherman, E. S. and Lippard, S. J.** (1987) Structural aspects of platinum anti-cancer drug interactions with DNA. *Chem. Rev.* **87**: 1153-1181.
- Snedecor, G. W. and Cochran, W. G.** (1976): *Statistical Methods* 6th Edition. Iowa State Univ. Press, Ames, Iowa.
- Verniorken, J., Van der Vijgh, W., Klein, L., Hart, A., Gall, H. and Pinedo, H.** (1984) Pharmacokinetics of free and total platinum species after short-term infusion of Cisplatin. *Cancer Treat. Rep.* **68**: 505-513.
- Ward, B. G., Harvey, V. J. and Shepherd, J. H.** (1982) Pregnancy after treatment of endodermal sinus tumour: case report with 5 years survival. *Br. J. Obstet Gynaecol.* **89**: 769-770.
- Zamble, D. B., Mu, D., Reardon, J. T., Sancar, A. and Lippard, S. J.** (1996) Repair of cisplatin-DNA adducts by the mammalian excision nuclease. *Biochemistry* **35**: 10004-10013.

(Received 02/02/2000, in revised form 24/03/2001)